

# Short Tandem Repeat Loci

Workshop

Phoenix, AZ

October 4, 2004

# DNA TYPING METHODS

- PCR-based
  - Faster results
  - Less labor-intensive
  - 1 ng DNA
  - Degraded DNA

# DNA TYPING METHODS

- D1 S80
  - single locus
  - preferential amp.
- DQA1 / PM
  - interpretational difficulties

- PCR-based
- Faster results
- Less labor-intensive
- 1 ng DNA
- Degraded DNA



Need for new typing method

Repetitive DNA contains sequences  
that occur more than once

# REPETITIVE DNA

(20-30% of human genome)

→ Interspersed:

→ Single repeats scattered throughout the genome

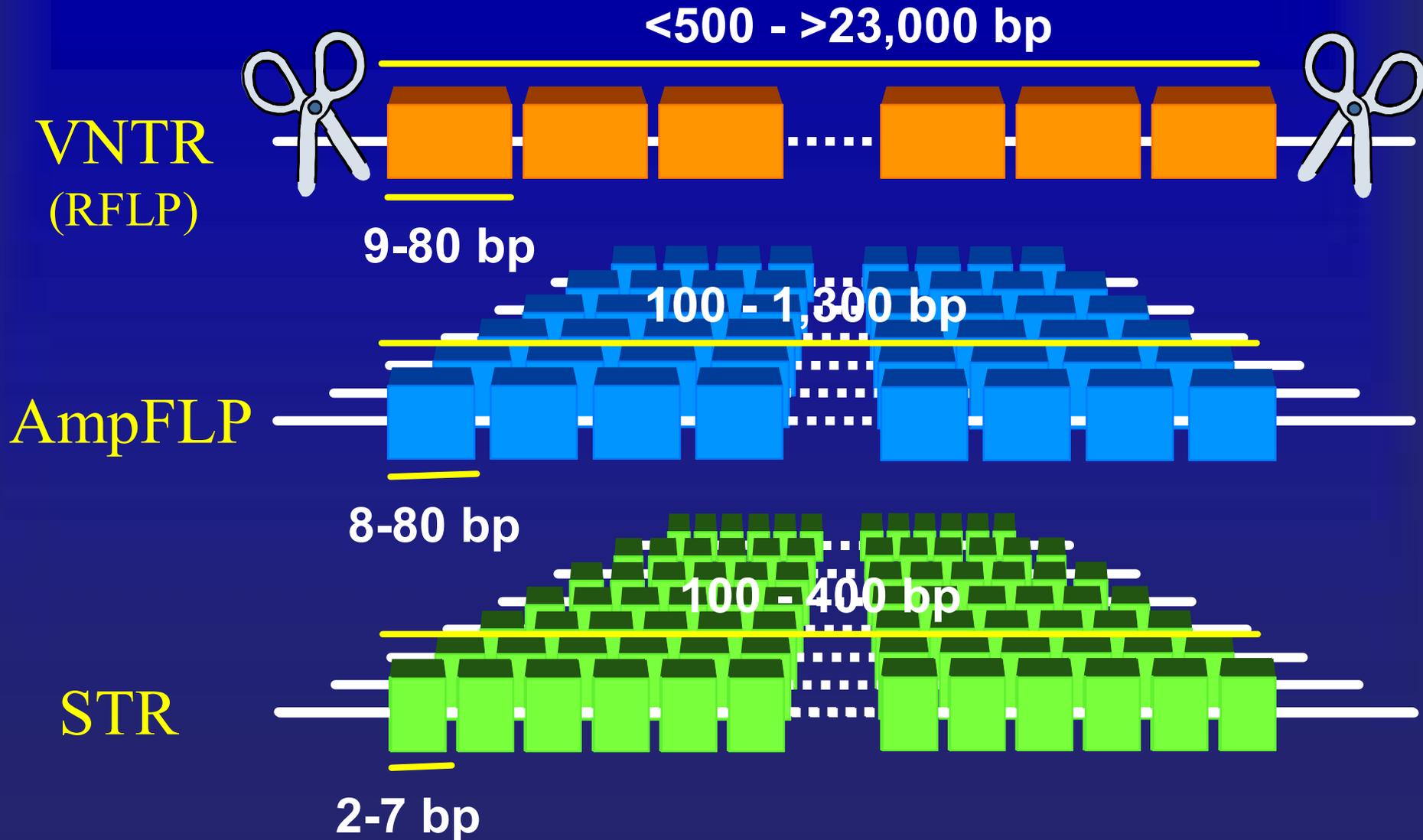
→ 10% of genome

→ Tandemly Repeated:

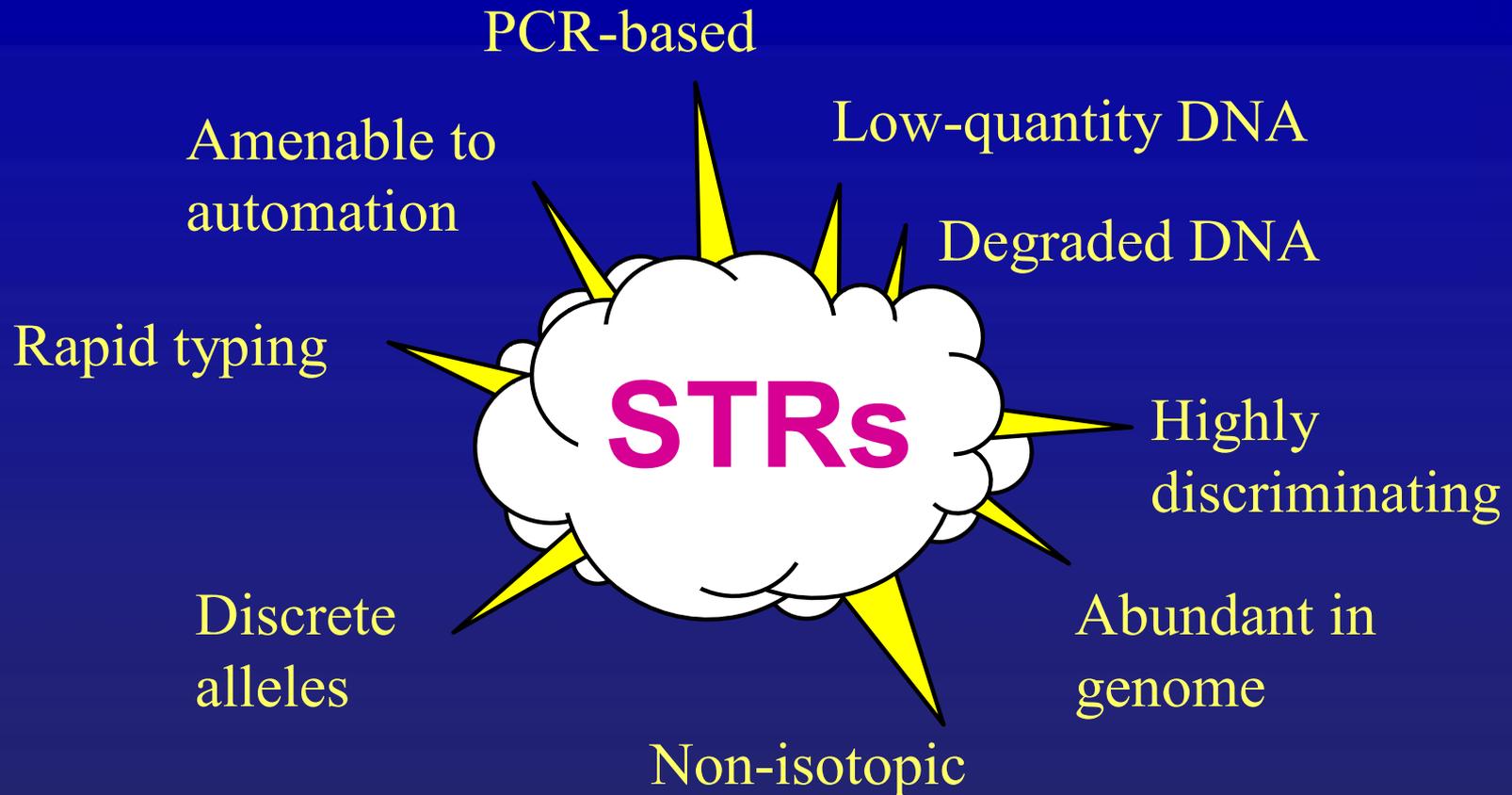
→ Head-to-tail tandem repetition of a common sequence

→ 15-20% of genome

# Repeat Polymorphisms



# Short Tandem Repeats



# SHORT TANDEM REPEATS

- Arrays of short repeats (2-7 bp) that are repeated several times in tandem
- >30,000 in the human genome
- One every 10 kb

# LENGTH OF REPEAT UNIT

2

Dinucleotides:

CA

3

Trinucleotides:

AAT

4

Tetranucleotides:

AGAT

5

Pentanucleotides:

AATGT

# GENERAL CRITERIA FOR SELECTING STR LOCI

2

Dinucleotide repeats:

Plentiful, but too much stutter

4

Tetranucleotide repeats:

Demonstrate less stutter

5

Pentanucleotide repeats:

Less stutter, but greater bp range

# GENERAL CRITERIA FOR SELECTING STR LOCI

4

Tetranucleotide repeats:

PD > 0.85

Size - 100-350 bps

Allele range < 60 bps

Low stutter

# TETRANUCLEOTIDE REPEATS

are used in forensics

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- Undesired PCR products are greater with di- and trinucleotide repeats
- Some trinucleotides are associated with diseases
- Alleles are more readily resolved
  - 4 bp, not 2 or 3 bp

# SHORT TANDEM REPEATS:

## Tetramers

AGAT    AGAT    AGAT    AGAT

AAAG    AAAG    AAAG    AAAG

AATG    AATG    AATG    AATG

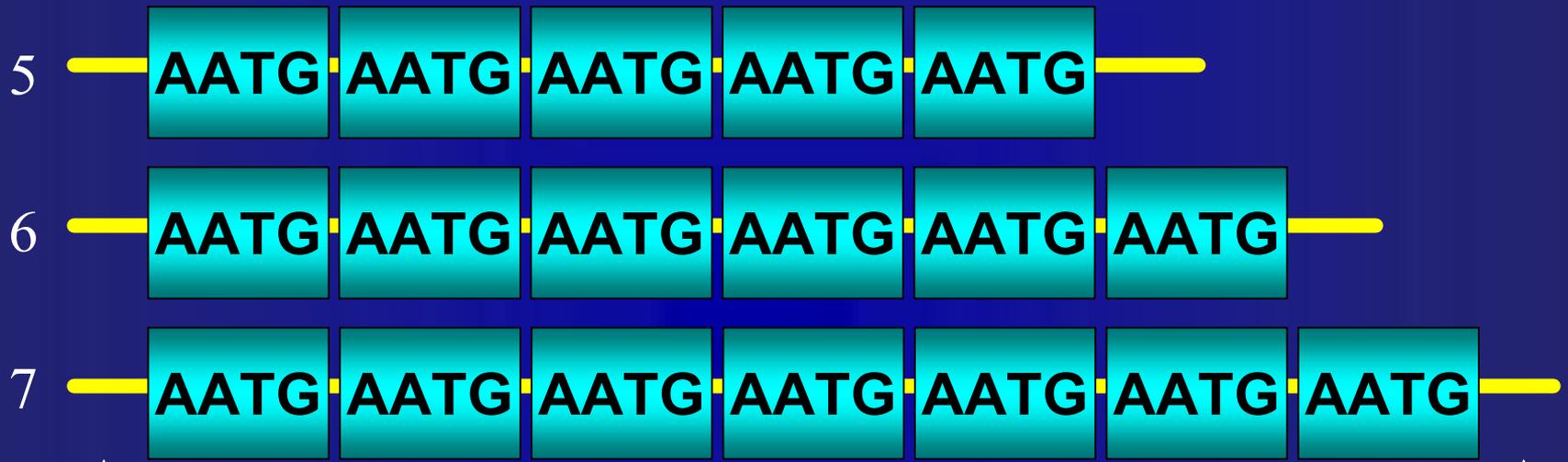
AAAT    AAAT    AAAT    AAAT

# STRs are POLYMORPHIC

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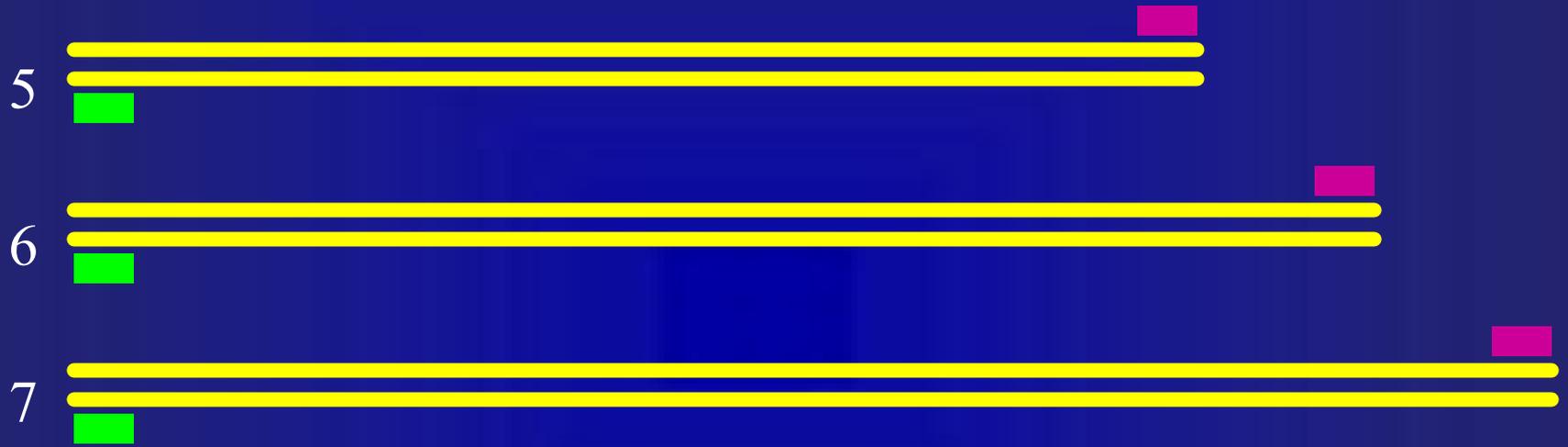
- Usually, variation in the number of tandem repeats that make up the array
- Alleles are designated by the number of repeats that they contain
- ★ Sequence variation can affect mobility and thus polymorphic nature - native vs denaturing conditions

Allele:



Flanking region of unique sequence

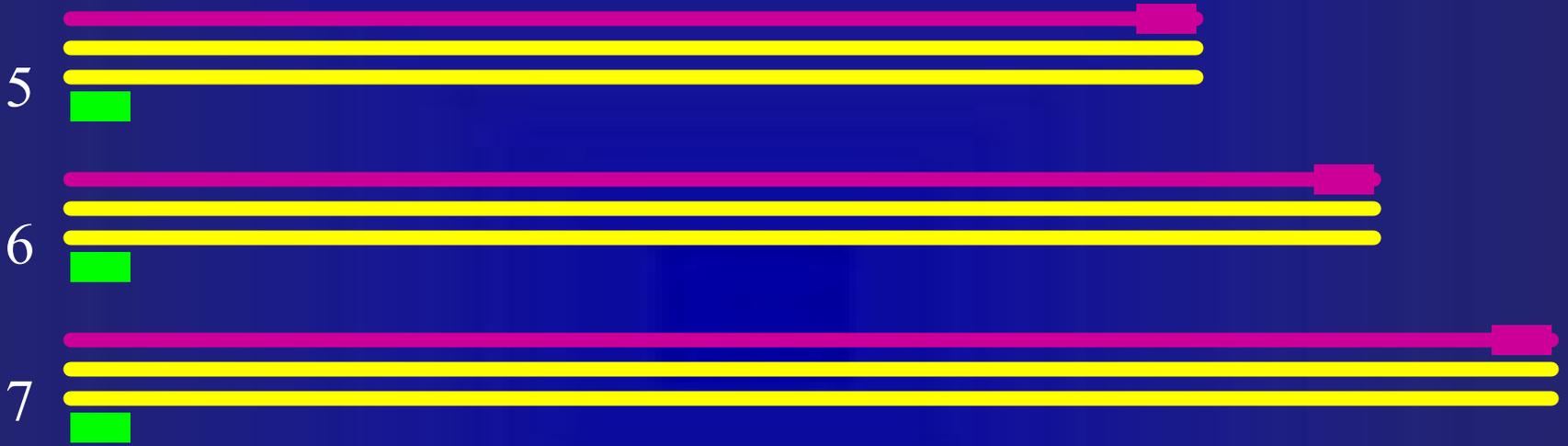
Allele:



.....PCR Primer Binding Sites.....

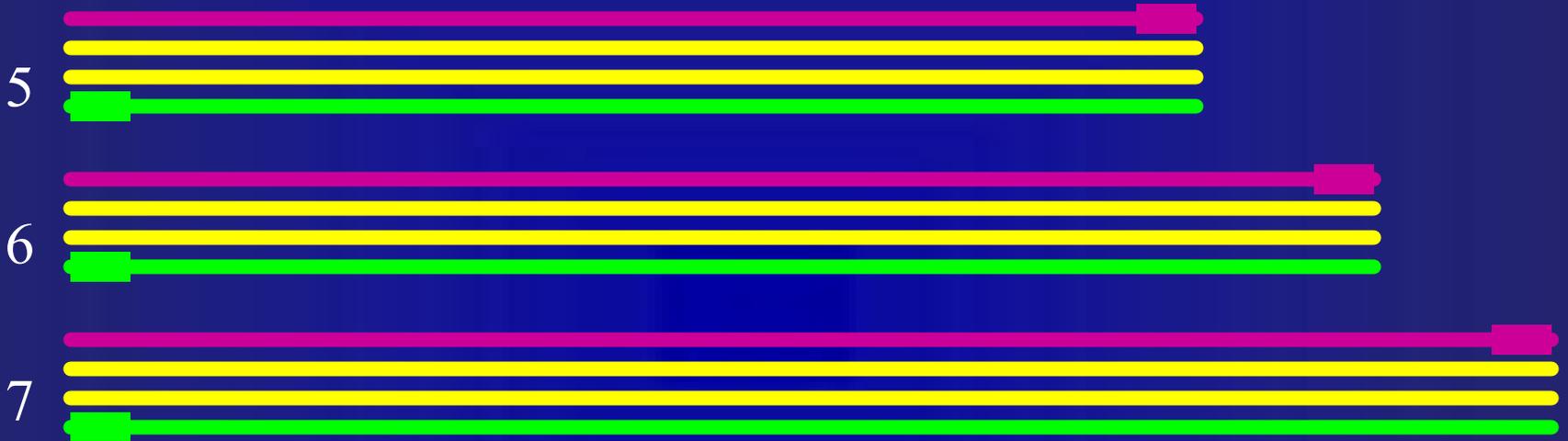


Allele:



.....Polymerase Chain Reaction.....

Allele:



.....Polymerase Chain Reaction.....

# TH01, a simple locus

→ (  AATG )<sub>5-11</sub>

**Operational  
sizing**

Allele:

5		169 bp*
6		173 bp
7		177 bp
8		181 bp
9		185 bp
10		189 bp
11		193 bp

\*AmpF1STR

# STR LOCUS TYPES

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- SIMPLE

- TH01: (AATG)<sub>5-11</sub>

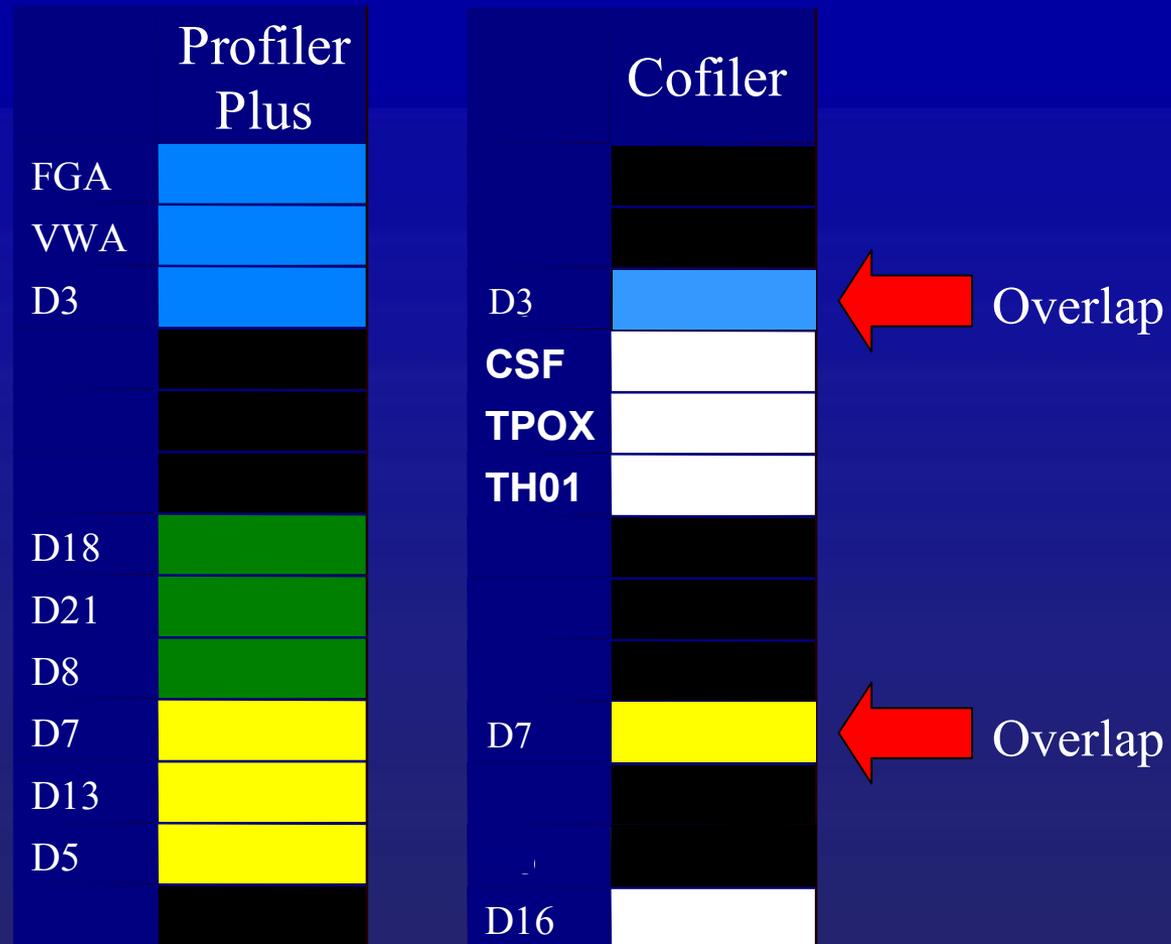
- COMPOUND

- VWA: TCTA (TCTG)<sub>3-4</sub>(TCTA)<sub>n</sub>

- COMPLEX

- D21S11:(TCTA)<sub>4-6</sub>(TCTG)<sub>5-6</sub>(TCTA)<sub>3</sub>TA  
(TCTA)<sub>3</sub>TCA(TCTA)<sub>2</sub>TCCATA  
(TCTA)<sub>8-16</sub>TC

# 13 Core Loci in 2 Amplifications



# 13 Core Loci Plus 2 more STR Loci in 1 Amplification

Penta D
D18S51
D21S11
THO1
D3S1358

FGA
TPOX
D8S1179
VWA
Amelogenin

Penta C
CSF1PO
D16S539
D7S820
D13S317
D5S818

**FGA**

Intron 3 of fibrinogen  
alpha chain

4q28

$(TTTC)_3$  TTTT TTCT  $(CTTT)_n$  CTCC  $(TTCC)_2$

**VWA**

Intron 40 of von  
Wildebrand factor

12p12-12pter

TCTA  $(TCTG)_{3-4}$   $(TCTA)_n$

**D3**<sub>S1358</sub>

Anonymous

3p

TCTA  $(TCTG)_{1-3}$   $(TCTA)_n$

**D18<sub>S51</sub>**

**Anonymous**

**18q21.3**

**(AGAA)<sub>n</sub>**

**D21<sub>S11</sub>**

**Anonymous**

**21**

**(TCTA)<sub>n</sub> (TCTG)<sub>n</sub> [(TCTA)<sub>3</sub>TA (TCTA)<sub>3</sub> TCA  
(TCTA)<sub>2</sub> TCCA TA] (TCTA)<sub>n</sub>**

**D8<sub>S1179</sub>**

**Anonymous**

**8**

**(TCTR)<sub>n</sub>**

**[R = A or G]**

**CSF<sub>1</sub>PO**

CSF-1 Receptor

**5q33.3-34**

**(AGAT)<sub>n</sub>**

**TPOX**

Intron 20 of thyroid  
peroxidase

**2p23-2pter**

**(AATG)<sub>n</sub>**

**TH01**

Intron 1 of tyrosine  
hydroxylase

**11p15.5**

**(AATG)<sub>n</sub>**

**D5**<sub>S818</sub>

**Anonymous**

**5q21-31**

**(AGAT)**<sub>n</sub>

**D13**<sub>S317</sub>

**Anonymous**

**13q22-31**

**(GATA)**<sub>n</sub>

**D7**<sub>S820</sub>

**Anonymous**

**7q**

**(GATA)**<sub>n</sub>

**D16S539**

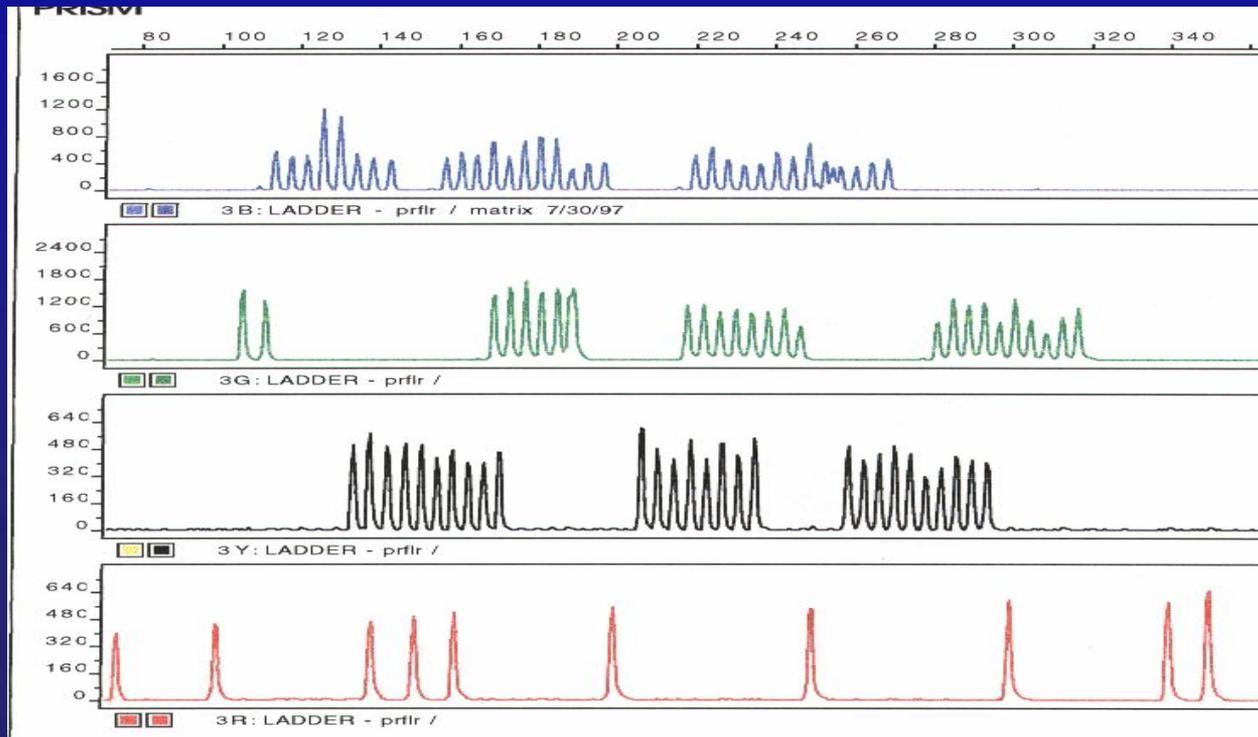
**Anonymous**

**16q24-qter**

**(AGAT)**

# ALLELIC LADDERS

Used to designate the alleles when genotyping an unknown sample



# ALLELIC LADDERS

- Simple Ladder is made with alleles that are 4-bp apart and that are within the range of common alleles
- Some variants may be included in the ladder (e.g., TH01 9.3)
- *Not all* observed variants are included in the ladder

# “OFF-LADDER” VARIANTS

Off-ladder (OL) alleles fail to align with the incremental 4-bp ladder or reside above or below ladder

To classify the variant allele:

Operationally defined nomenclature

Number of repeats,  
followed by a decimal point  
number of nucleotides of  
partial repeat

Above or below ladder  
use > or < highest or  
lowest allele in ladder

# “OFF-LADDER” VARIANTS

TH01

9

( AATG )<sub>9</sub>

9.3

( AATG )<sub>6</sub>

**ATG**

( AATG )<sub>3</sub>

10

( AATG )<sub>10</sub>

# “OFF-LADDER” VARIANTS

## TH01

9 (AATG)<sub>9</sub>

9.3 (AATG)<sub>6</sub> ATG (AATG)<sub>3</sub>

10 (AATG)<sub>10</sub>

## VWA

16 (TCTA)<sub>2</sub> (TCTG)<sub>4</sub> (TCTA)<sub>10</sub>

16.2 (TCTA)<sub>2</sub> (TCTG)<sub>4</sub> TA (TCTA)<sub>10</sub>

# Designing STR Systems

- Primers
- Test primers for compatibility:  
specificity, balance, primer dimer,  
full A addition
- Test PCR components  
MgCl<sub>2</sub>, KCl, dNTPs, template
- Thermal cycler conditions
- Post-PCR analytical procedures  
fluors, electrophoresis, detection  
systems, interpretation

# STR Primer Design

- No size overlap for loci with same fluor
- Similar  $T_m$  for primers
- Avoid hairpin structures
- Minimal complementary binding at 3' end
- Maximal 3' A addition
- Test multiplex at low  $T_{anneal}$  for specificity

# Specificity



# AmpliTaq Gold

- Hot start
- Simplifies PCR set up
- Reduces primer design requirements
- Decreases primer-dimer formation
- Facilitates multiplexing
- Automation

60°C

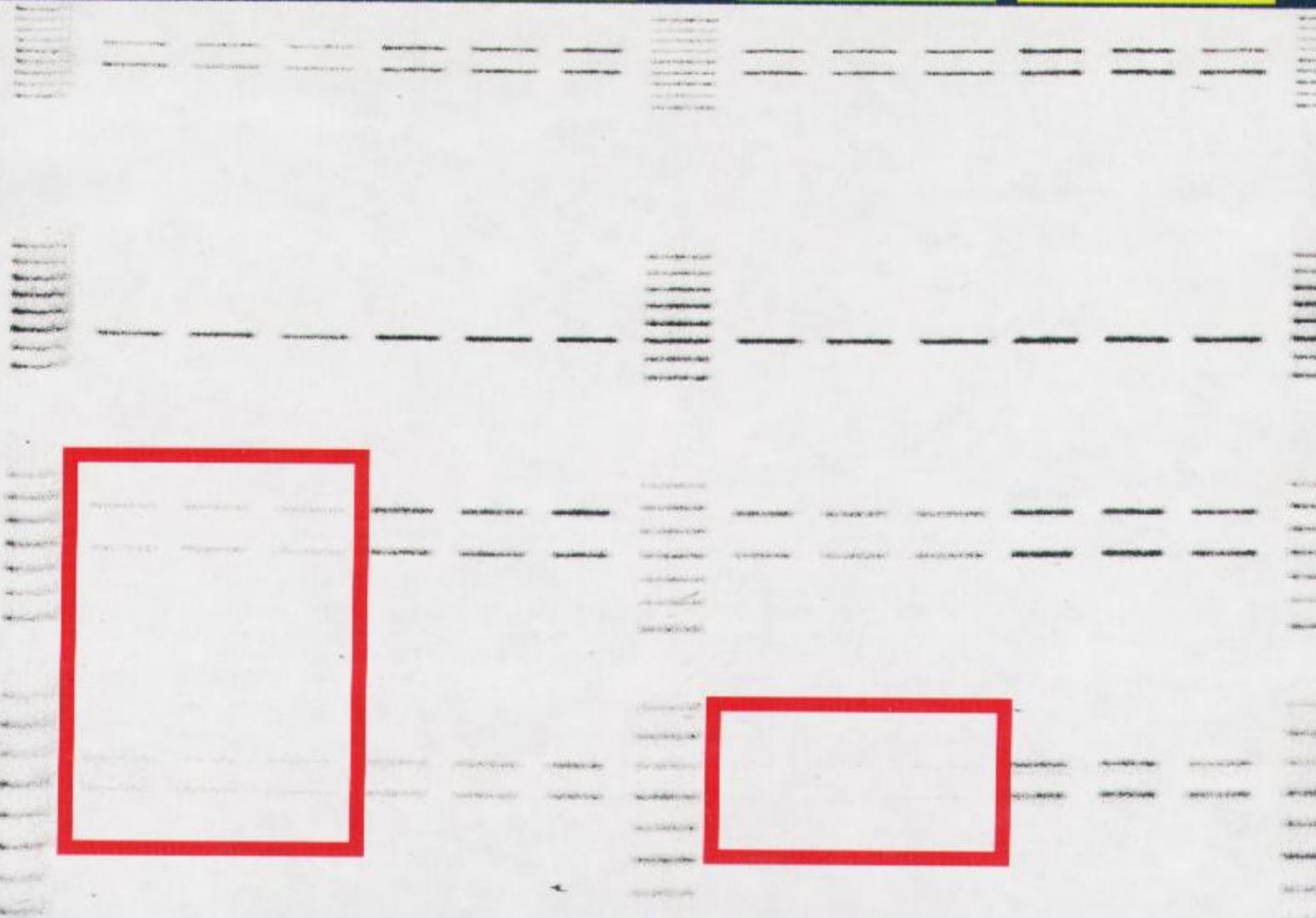
64°C

Taq

Gold

Taq

Gold

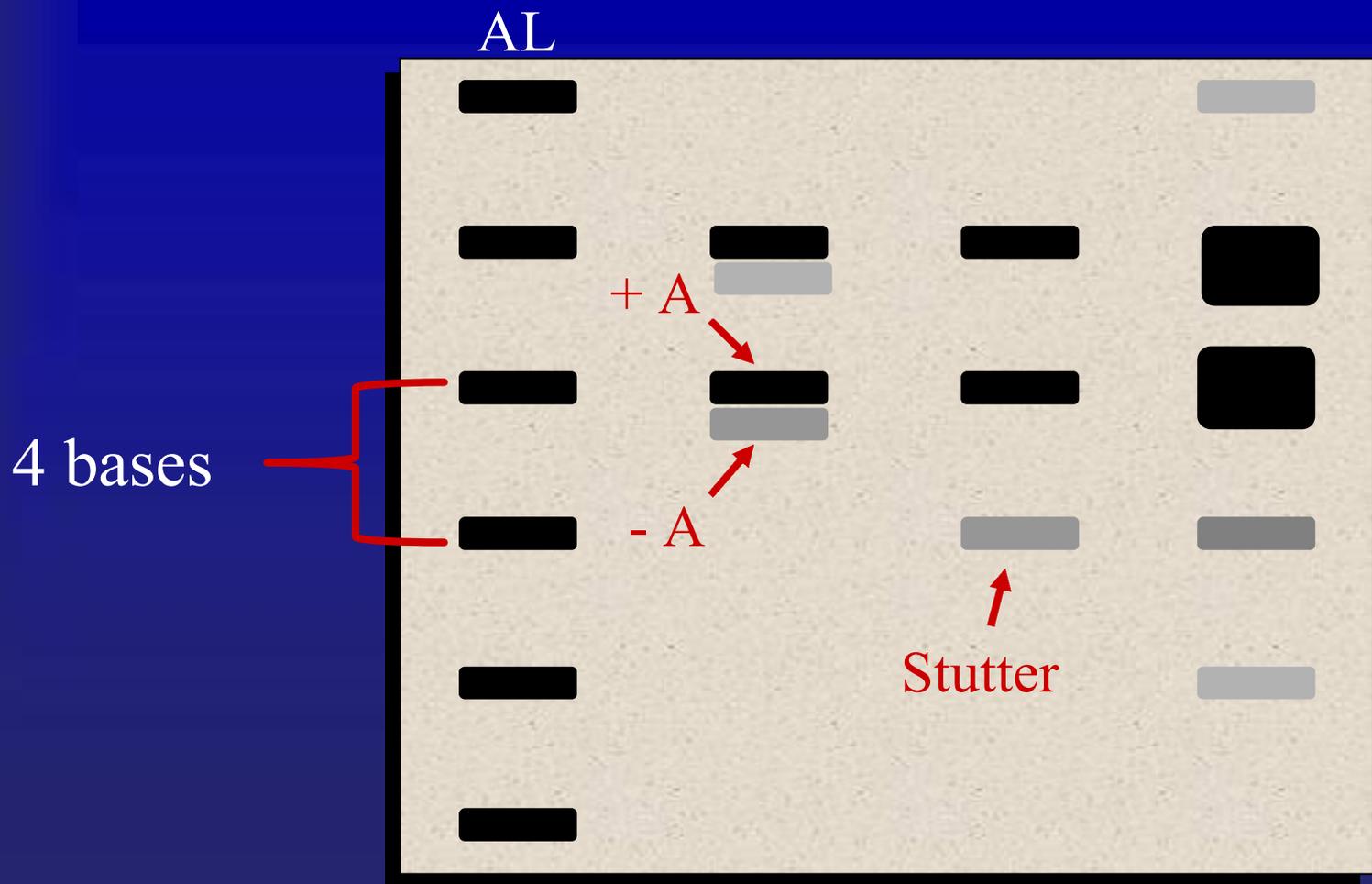


Undesired!  
But inherent.

# UNDESIRE PCR PRODUCTS

“Minus A” or “Plus A”  
Stutter

# The difference between “Minus A” and Stutter



# Non-Templated Nucleotide Addition (“Plus A”)

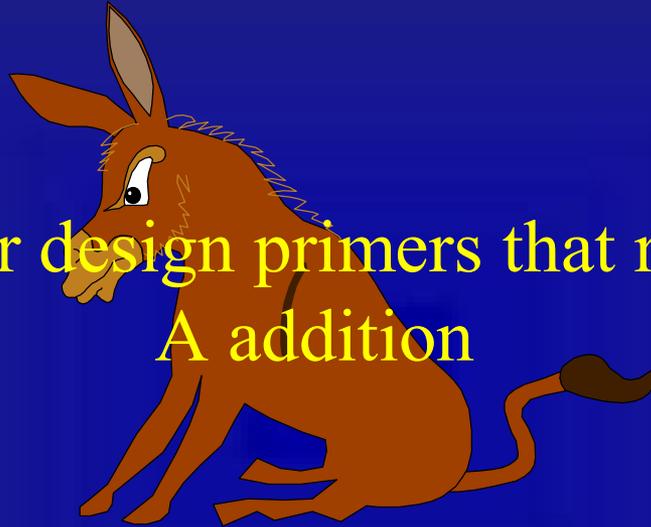
- Many DNA polymerases add an extra nucleotide to the blunt end of a DNA product *without* the use of a template
- Usually A is added
- The resulting band/peak is 1-bp larger in size than that predicted from the DNA sequence and primer locations

# Non-Templated Nucleotide Addition (“Plus A”)



There are two ways to deal with +/-A

Either design primers that resist  
A addition



Or drive the reaction towards  
addition of A



If you can't beat 'em,  
join 'em

One cannot effectively prevent +A.  
However, one can increase +A  
through primer design...

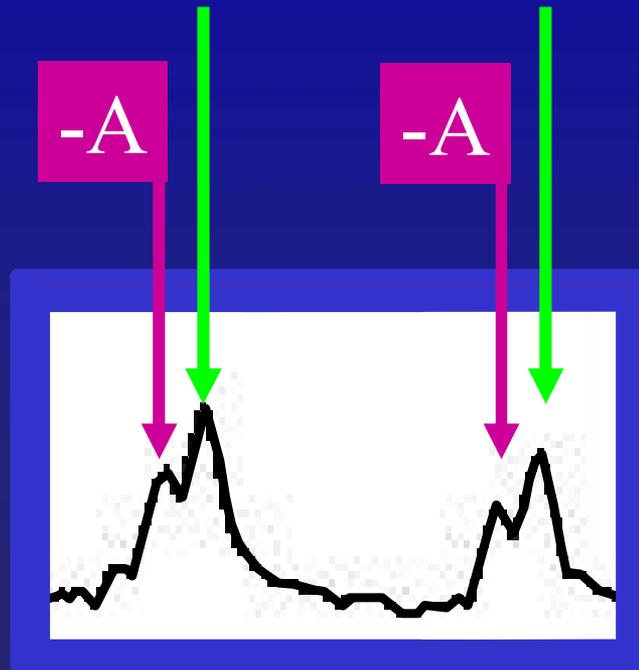
“Plus A” is the desired  
product.

Ladder bands / peaks are +A!

Inefficient adenylation results in 2 bands  
or peaks  
(or odd-shaped peaks)

Allele 1, +A

Allele 2, +A

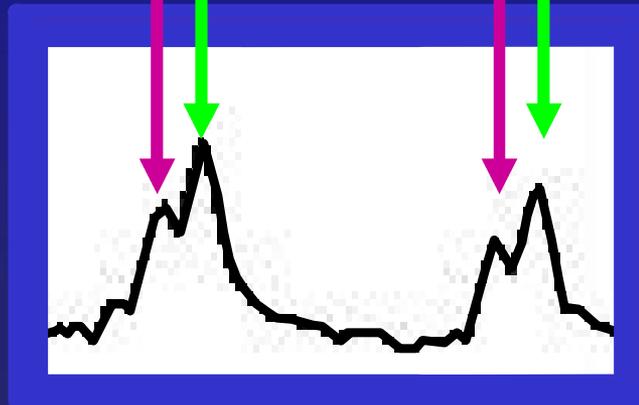


# Inefficient adenylation results in 2 bands or peaks (or odd-shaped peaks)

Reduced Peak Height

OL Allele?

Or -A?



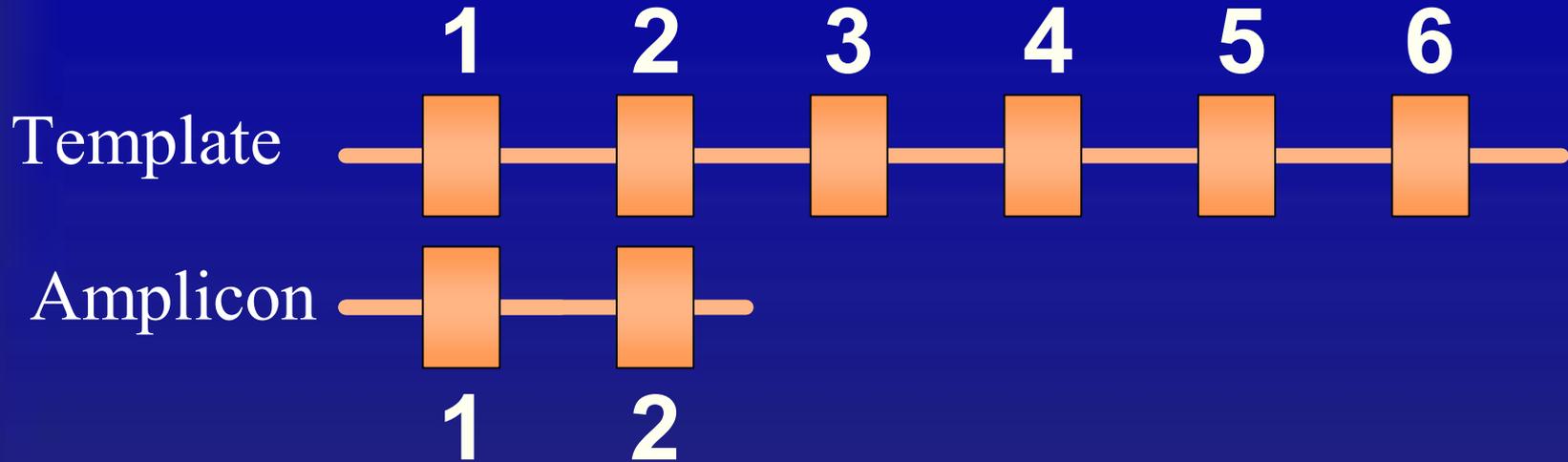
# ADDRESSING “MINUS A” PEAKS

- Repeat final extension step with additional enzyme
- Use knowledge of adenylation efficiency at a given locus
- Use database information to aid in interpretation

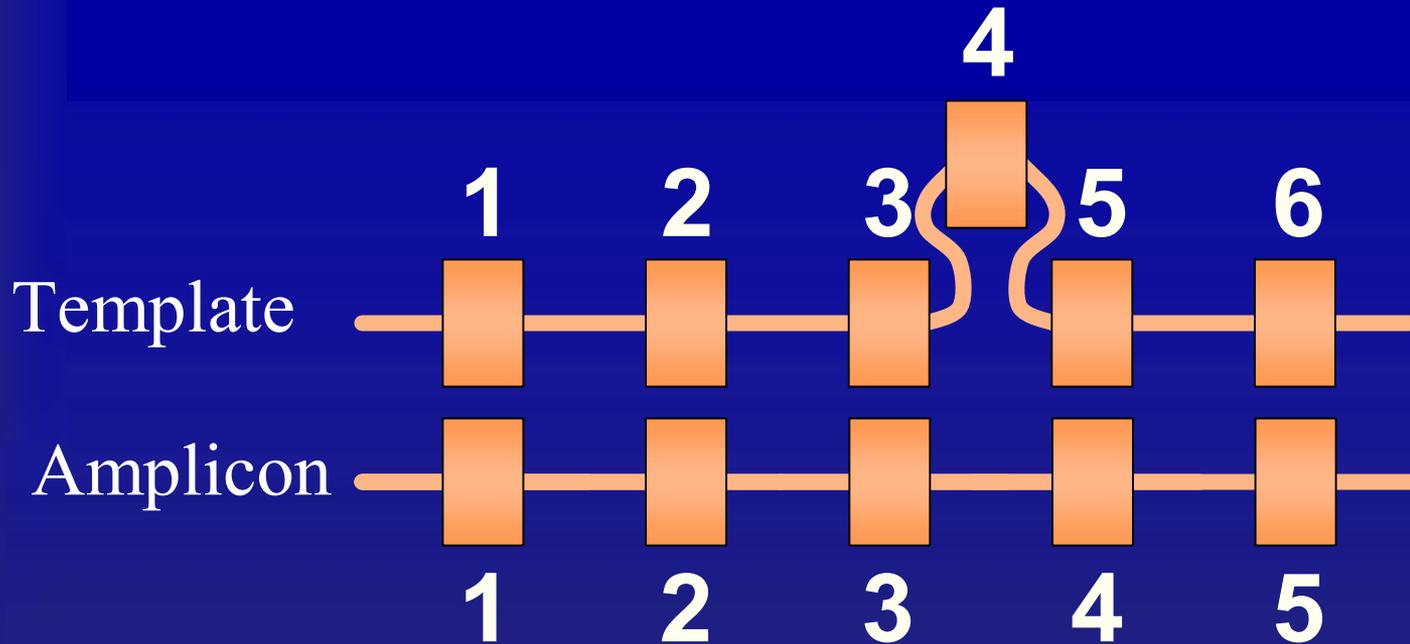
# STUTTER

- Characteristic of repetitive DNA amplification
- Tetramers have less stutter than dinucleotide repeats
- Usually one repeat unit less in size
- Increases with magnesium
- 0-14% of allele

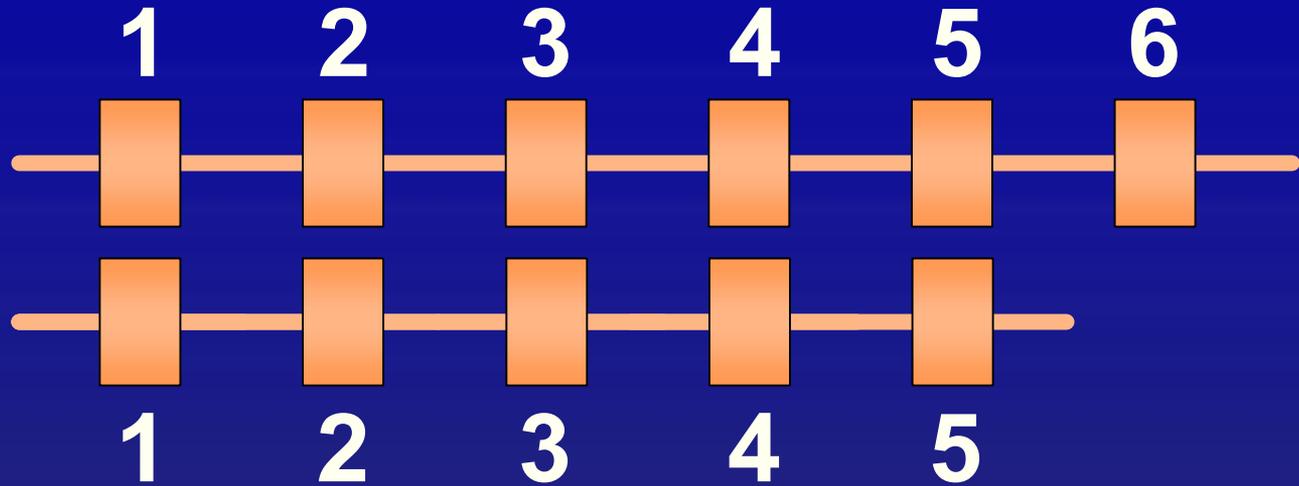
# HOW STUTTER OCCURS



# HOW STUTTER OCCURS



# HOW STUTTER OCCURS





6

5

TRUE ALLELE PREDOMINATES

# Minimize stutter by working within the recommended peak height range

5-fold more product

Actual % stutter is the same

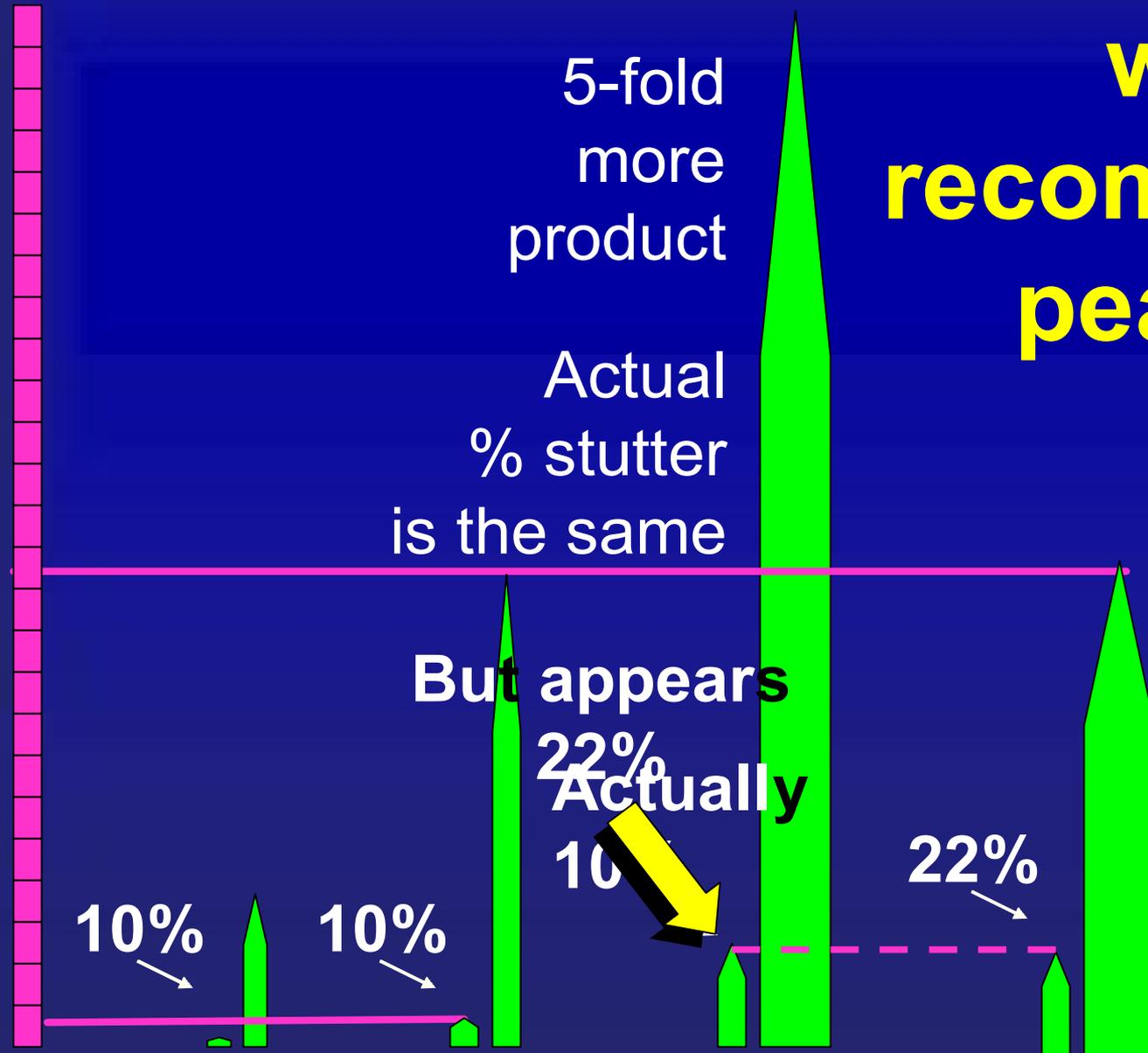
But appears 22%  
Actually 10%

Threshold of detection is exceeded for main peak ...but not for stutter peak

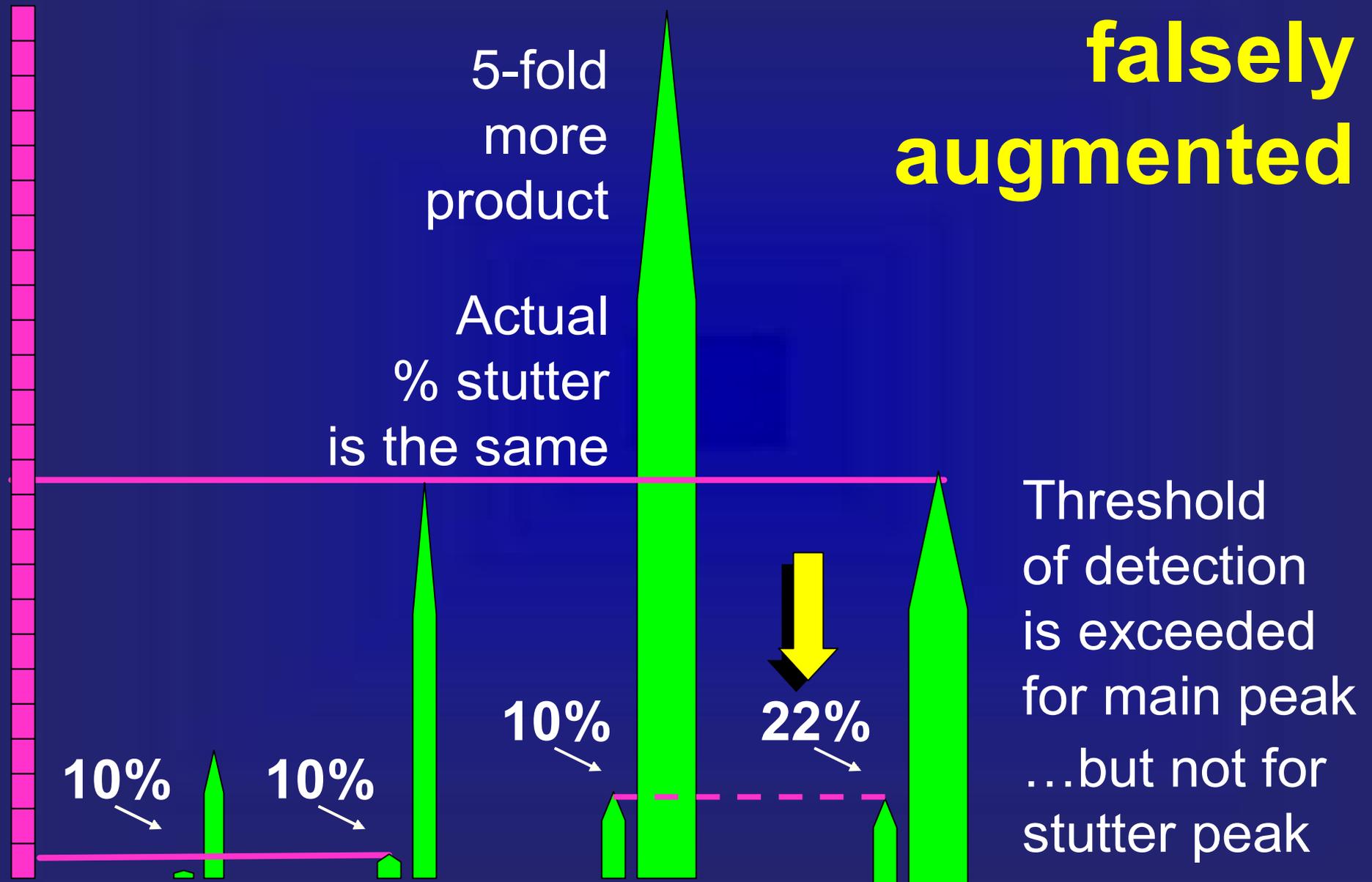
10%

10%

22%



# Percent stutter is falsely augmented



# STUTTER TRENDS

- Different loci exhibit different degrees of stutter
- Loci with “compound” or “complex” structures exhibit less stutter than those with “simple” structure (e.g., VWA)
- Also variant alleles at a locus may have less stutter (e.g., VWA)

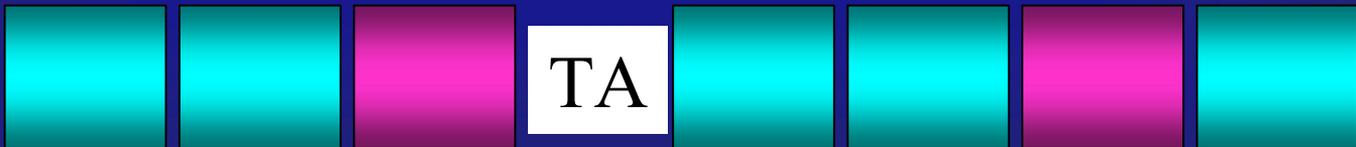
Simple:



Compound:



Complex:



# Allele 16

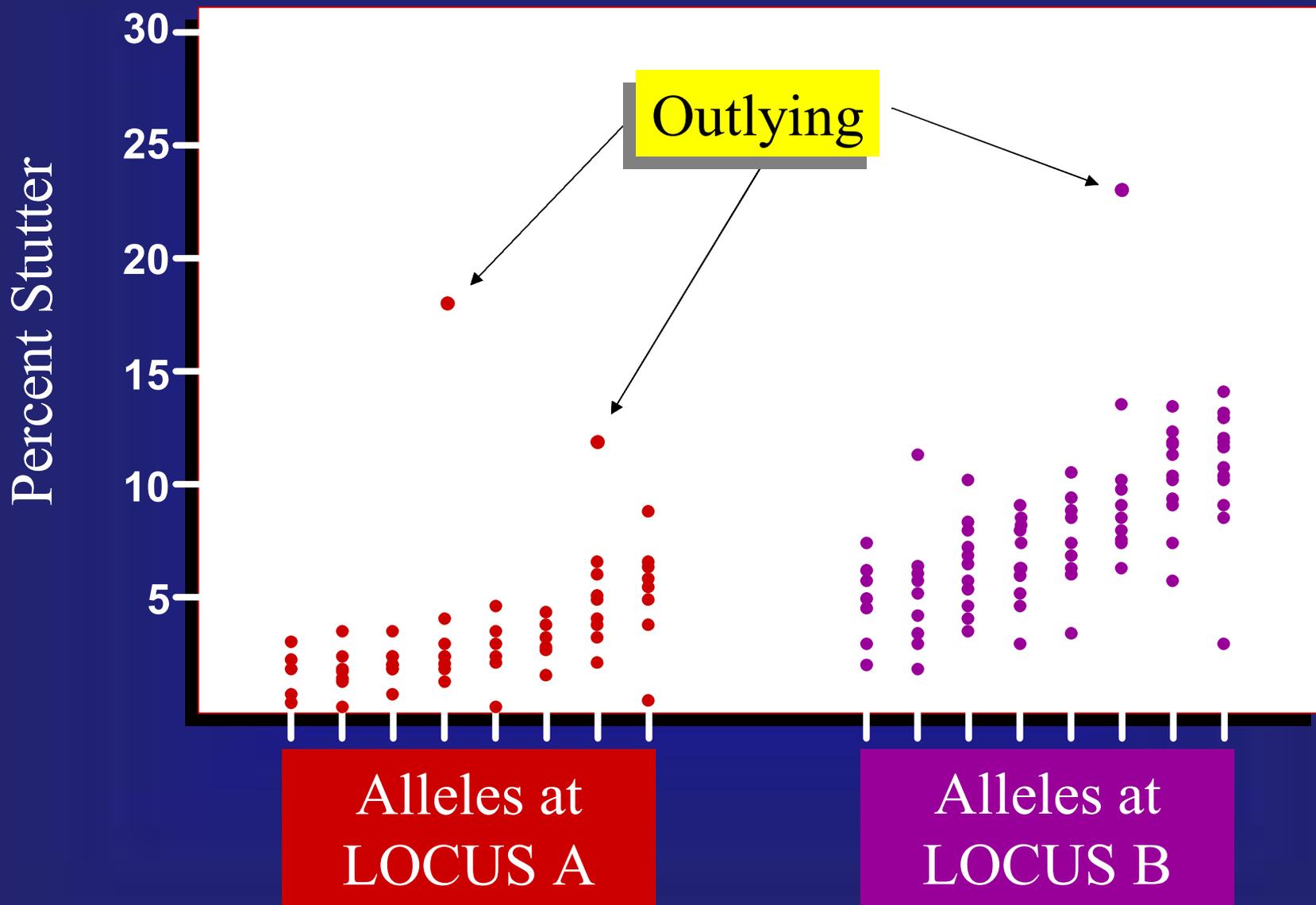
16 ( TCTA )<sub>2</sub> ( TCTG )<sub>4</sub> ( TCTA )<sub>10</sub>

16 ( TCTA )<sub>2</sub> ( TCTG )<sub>4</sub> ( TCTA )<sub>4</sub> ( TCTG ) ( TCTA )<sub>5</sub>

Less prone to stutter

# STUTTER TRENDS

- Different loci exhibit different degrees of stutter
- Loci with “compound” or “complex” structures exhibit less stutter than those with “simple” structure
- At any given locus, smaller alleles typically exhibit a lower percent stutter than larger alleles



# STUTTER TRENDS

- Different loci exhibit different degrees of stutter
- Loci with “compound” or “complex” structures exhibit less stutter than those with “simple” structure
- At any given locus, smaller alleles typically exhibit a lower percent stutter than larger alleles
- General guidelines

# The presence of stutter or -A is not necessarily a problem

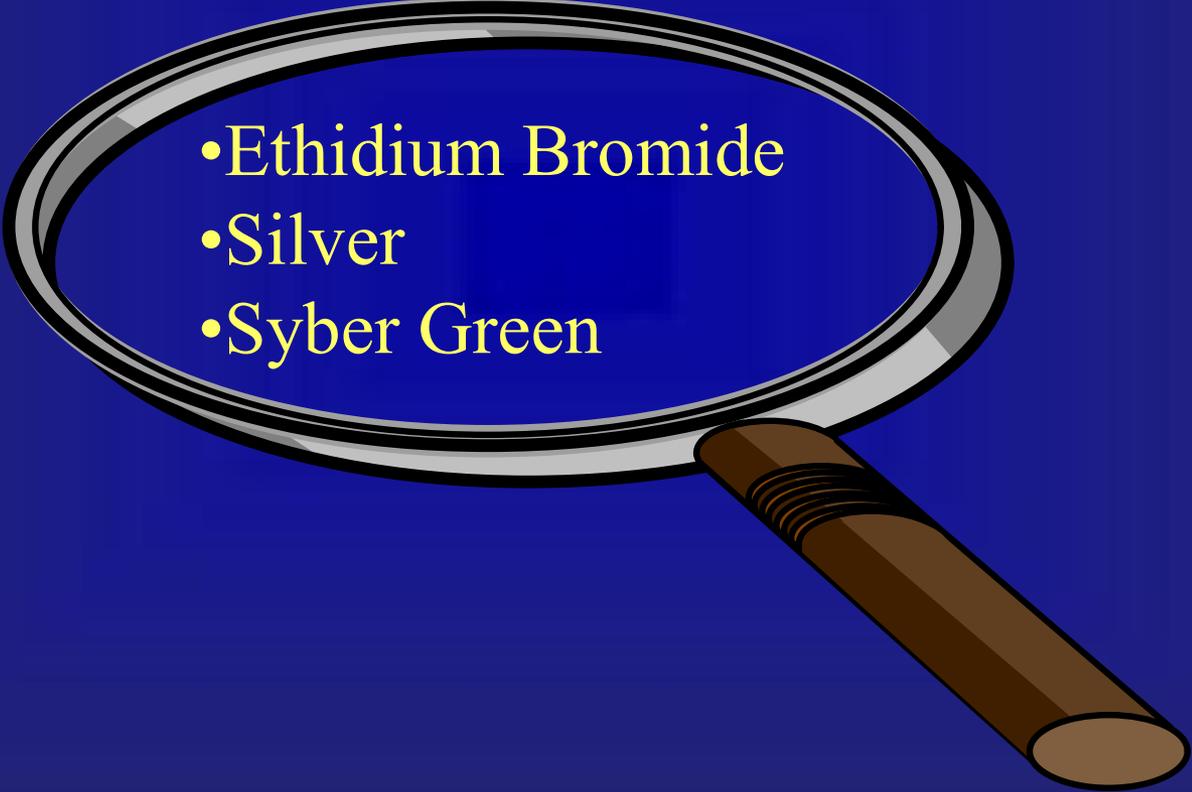
- Stutter is a feature of STR amplification
- Low-level stutter is manageable, when peaks are within recommended range
- Mixture interpretation
- Samples with -A can be extended
- If +A failure occurs, both alleles at a heterozygous locus tend to exhibit -A
- Adenylation failure does not preclude correct genotyping

# Resolution

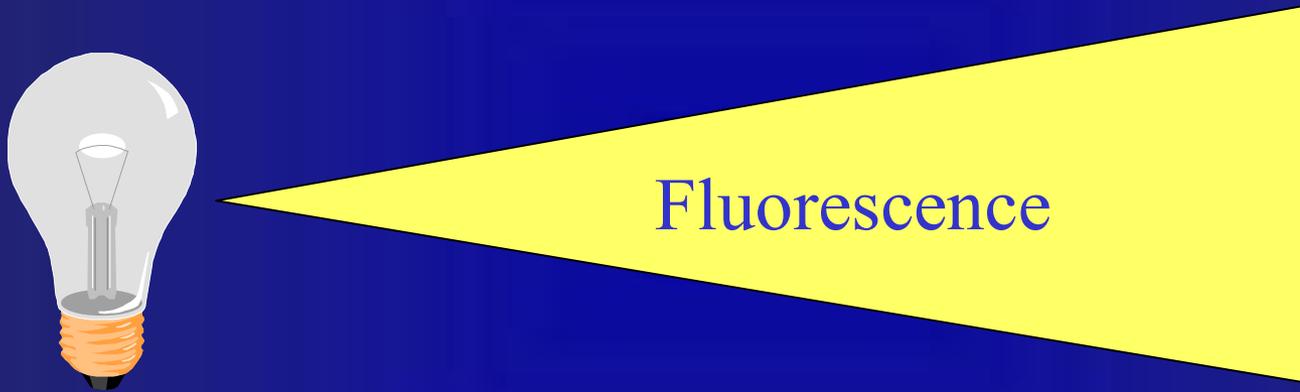
## Typing

- SA-32 Gel Apparatus, FMBIO
  - Scan gel after run
- ABI Prism 377, 310, 3100, 3700,...
  - Detection window

# Detection Methods

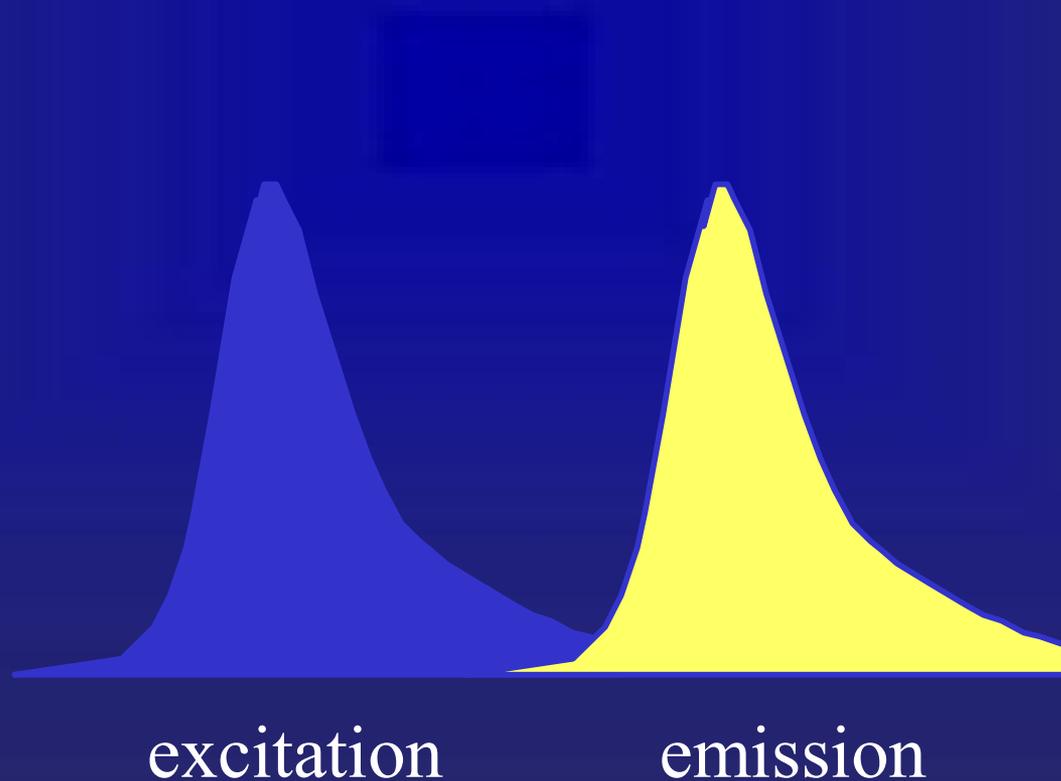
- 
- Ethidium Bromide
  - Silver
  - Syber Green

# Detection Methods



Label one of the primers for each locus

Fluorescence results when a fluorophore or fluorescent dye absorbs incident light and in response emits light at a different wavelength

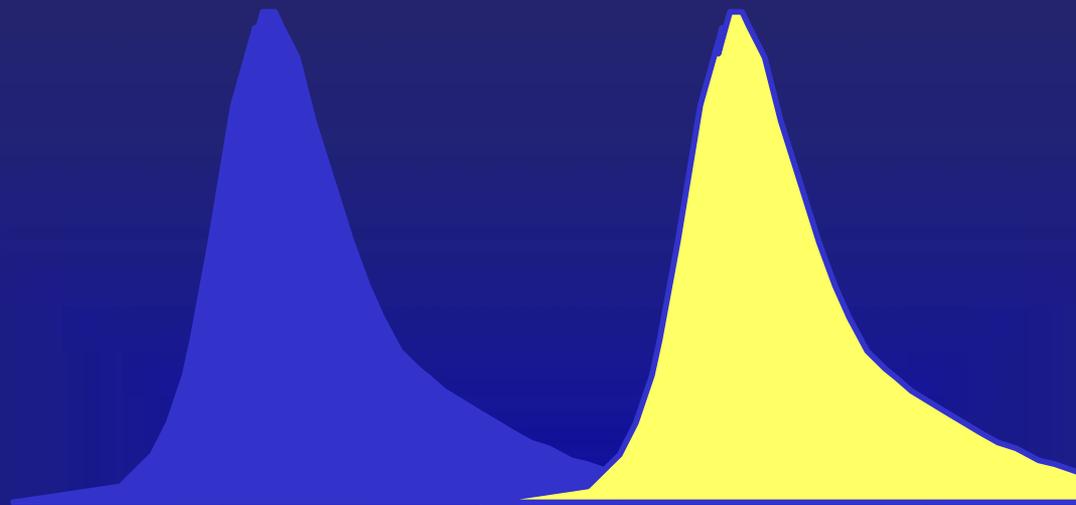


## Molar Extinction Coefficient is

a measure of a dye's ability to absorb light

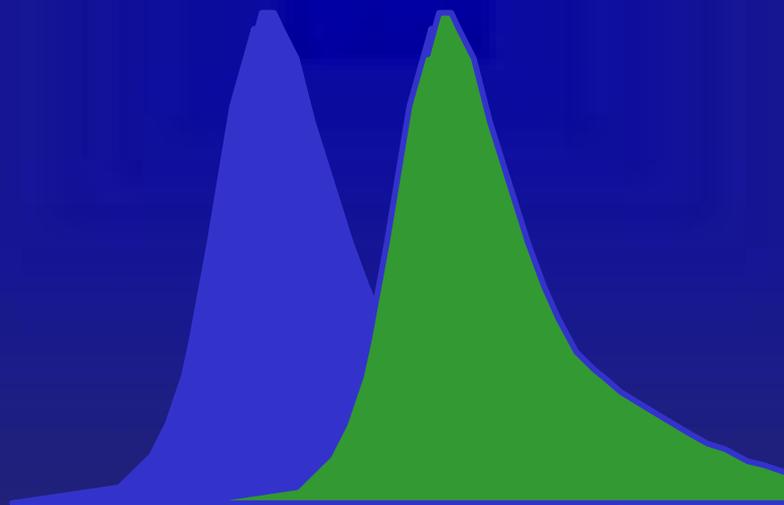
## Stokes Shift is

The difference in wavelength between the fluorescence excitation maximum and fluorescence emission maximum



excitation

emission



excitation

emission

## Spectral Criteria

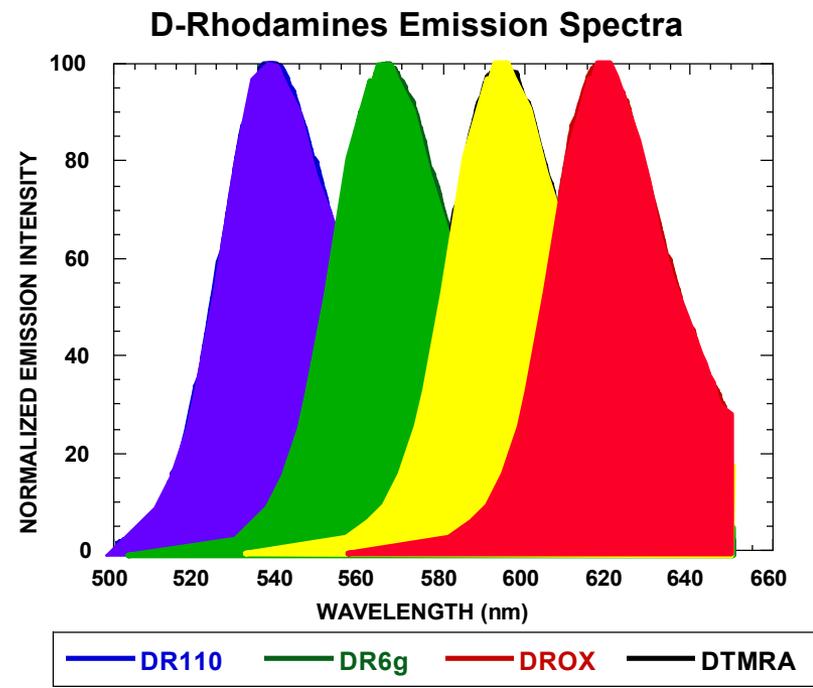
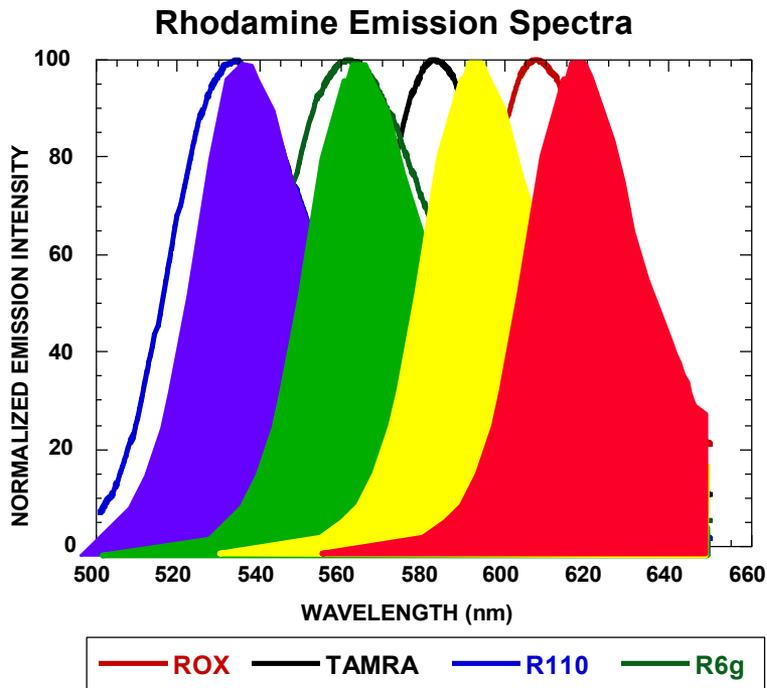
The dye's spectral excitation and spectral emission wavelengths should be compatible with excitation source and filter wavelengths

## Spectral Bandwidth

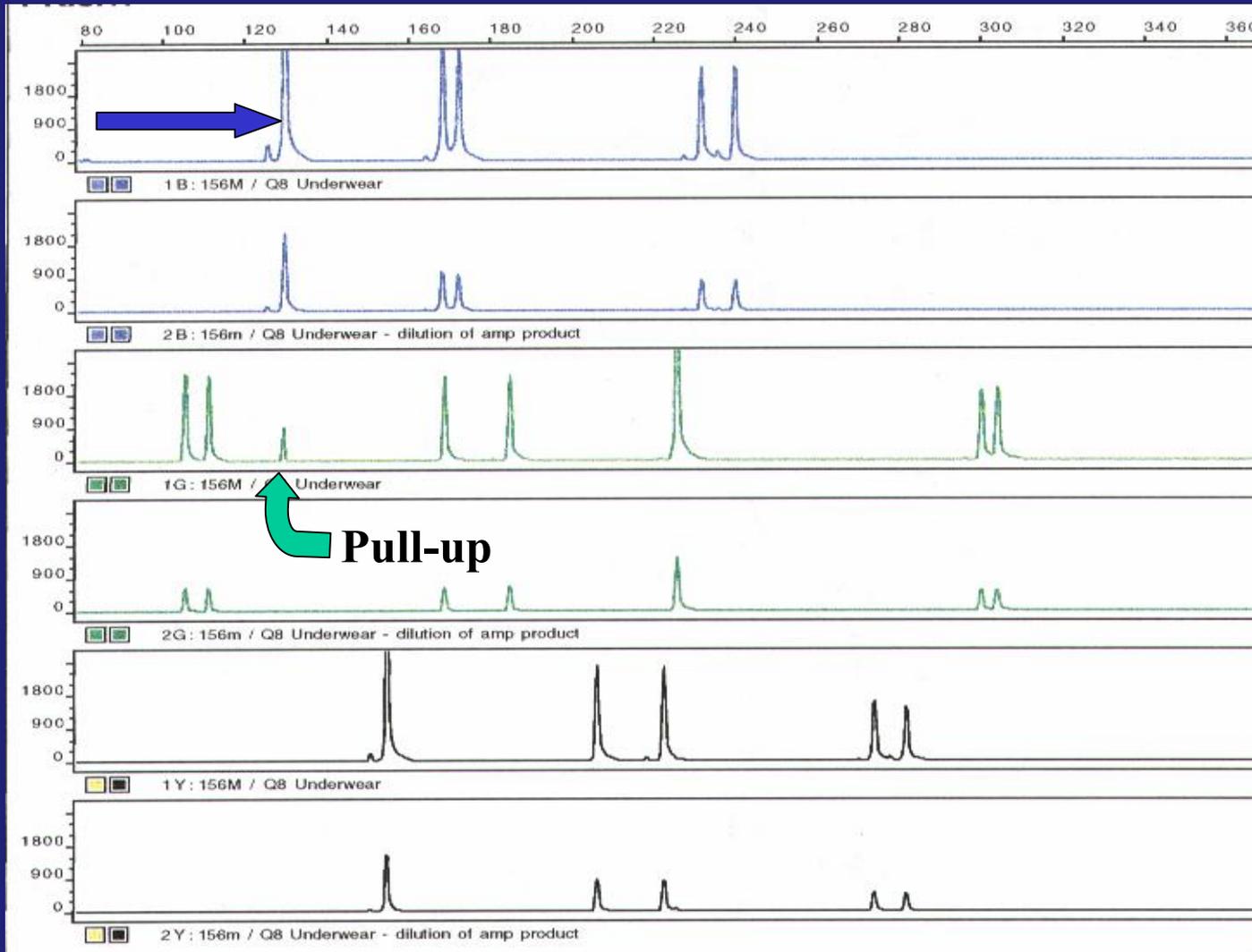
Spectral overlap and the ability of an instrument to distinguish one dye from another

**PULL-UP**

# Comparison of Dichlororhodamine and Rhodamine dye spectra



# Pull-up



Due to signal and spectral overlap

## Fluorescence Quantum Yield is

The efficiency with which the excited molecule is able to convert absorbed light to emitted light

**Dyes also can affect**  
Mobility of DNA molecules

# SUMMARY

- STRs used today in forensics generally are characterized by tandem repeats of 4(or 5)-bp units
- STR polymorphism is due to variation in the number of repeated units
- There are different repeat unit sequences (AATG, TCTA, CTTT, etc.)
- STR loci are either simple, compound or complex

# SUMMARY

- Alleles are designated by comparison with an allelic ladder
- Off-ladder variants of tetramers usually have 1, 2, or 3 extra bases within the repeat array
- Off-ladder variants also can occur above and below the extremities of the ladder

# SUMMARY

- STR loci “associated with” genes are typically in non-coding regions (i.e., introns or flanking regions)
- Many STR loci are “anonymous,” or not currently known to reside within a known gene

# SUMMARY

- STRs are amplified by the PCR. PCR primers bind to unique sequences that flank the repeat region
- Alleles that align with the allelic ladder have “plus A”
- “Minus A” may occur at some loci but is easily addressed

# SUMMARY

- Stutter is inherent to STR (tetramers) amplification of some loci
- Loci with a simple repeat structure tend to stutter more
- Loci vary in the extent to which they exhibit stutter
- Small alleles show less stutter than larger alleles
- Stutter is generally predictable and general guidelines can be used for profile interpretation

# SUMMARY

- Interpreting electropherograms is facilitated, especially with software

THE END